

Figure 1: Molecular characterization of putative IP₃ receptor (IP₃R) from *Leishmania infantum*

(A) Schematic representation of macronuclear sequence of putative IP₃R gene *LinJ16_V30290* from *Leishmania infantum*.

The *LinJ16_V30290* gene (chromosome 16) is flanked upstream by gene *LinJ16_V30300* (putative proteasome 26S non-ATPase subunit 9) and downstream by gene *LinJ16_V30280* (conserved hypothetical protein). The *LinJ16_V30290* gene is intronless.

(B) Domain structure of the *Leishmania infantum* putative IP₃R.

Protein BLAST search (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) against reference proteins from mouse (*Mus musculus*) showed that the protein encoded by *LinJ16_V30290* had highest sequence similarity with IP₃R type 2 (IP₃R2) and Ryanodine receptor type 3 (RyR3) which were further used for global sequence similarity and identity analysis by the SIAS server (<http://imed.med.ucm.es/Tools/sias.html>). Such analysis revealed that the putative IP₃R from *Leishmania infantum* had a global sequence identity and similarity of 11.4% and 54.3% respectively with IP₃R2 and 5.02% and 9.46% respectively with RyR3.

Domain analysis was conducted using the InterProScan server (<http://www.ebi.ac.uk/Tools/pfa/iprscan/>) and transmembrane domains were determined by the TMHMM server (<http://www.cbs.dtu.dk/services/TMHMM/>). Results are presented relative to scale of the full-length amino acid (AA) sequence of the protein.

(C) Analysis of transmembrane domains of putative IP₃R from *Leishmania infantum* (left) and mouse IP₃R2 (right) using the TMHMM server (<http://www.cbs.dtu.dk/services/TMHMM/>).

(D) Hydrophobicity analysis of the transmembrane domains of putative IP₃R from *Leishmania infantum*.

Hydrophobicity plots reveal the presence of 5 transmembrane regions compared to 6 that are present in the mouse IP₃R2 (see Panel C (right)).

(E) Clustal W2.0.12 alignment of the amino acid sequence between transmembrane domains 4 and 5 of putative IP₃R from *Leishmania infantum* and the amino acid sequence between transmembrane domains 5 and 6 of mouse IP₃R2 and mouse RyR3. The selectivity filter of mouse IP₃R2 is shown as boxed text.

(F) Modeling of the putative IP₃R-ligand binding suppressor domain of *Leishmania infantum* using the EsysPred3D homology-modelling server (<http://www.fundp.ac.be/sciences/biologie/urbm/bioinfo/esypred/>). Structure of the IP₃R-ligand binding suppressor domain of mouse IP₃R type 1 (right), (ITPR1; <http://www.pdb.org/pdb/explore/explore.do?structureId=1XZZ>) (Bosanac *et al.*, 2005) and model of the *Leishmania infantum* (Lin; left) homologous region. Numbers indicate the amino acid residues used.

(G) Modeling of the putative IP₃R-ligand binding core from *Leishmania infantum* using the EsysPred3D homology-modelling server (<http://www.fundp.ac.be/sciences/biologie/urbm/bioinfo/esypred/>). Structure of the IP₃R-ligand binding core of mouse IP₃R type 1 (right), (ITPR1; <http://www.pdb.org/pdb/explore/explore.do?structureId=1N4K>) (Bosanac *et al.*, 2002) and model of the *Leishmania infantum* (Lin; left) homologous region. Numbers indicate the amino acid residues used. The arrow indicates the IP₃ binding site.

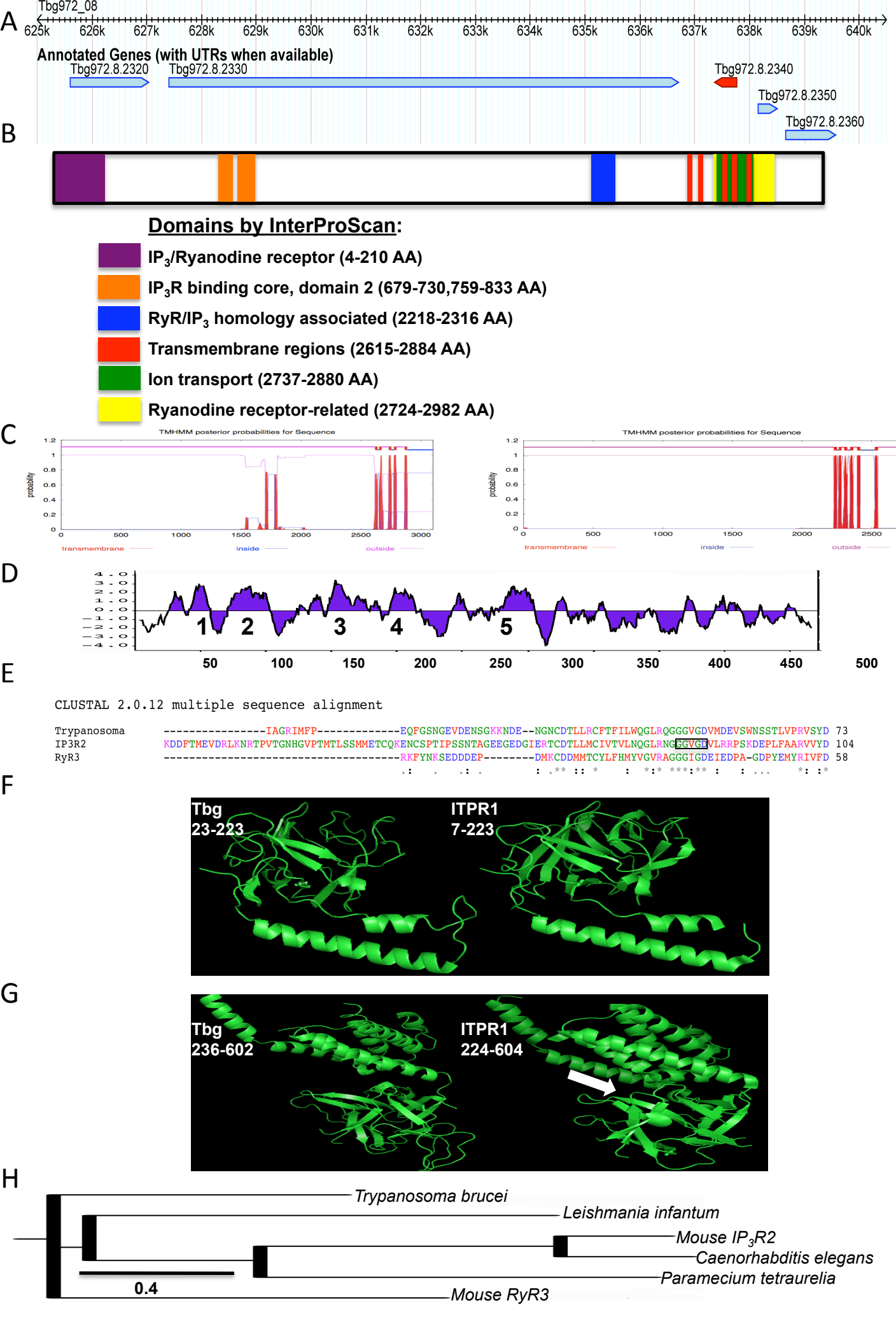


Figure 2: Molecular characterization of putative IP₃ receptor (IP₃R) from *Trypanosoma brucei*

(A) Schematic representation of macronuclear sequence of putative IP₃R gene *Tbg972.8.2330* from *Trypanosoma brucei*.

The *Tbg972.8.2330* gene (chromosome 8) is flanked upstream by gene *Tbg972.8.2320* (conserved hypothetical protein) and downstream by gene *Tbg972.8.2360* (putative RNA-binding protein RBP10). The *Tbg972.8.2330* gene is intronless.

(B) Domain structure of the *Trypanosoma brucei* putative IP₃R.

Protein BLAST search (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) against reference proteins from mouse (*Mus musculus*) showed that the protein encoded by *Tbg972.8.2330* had highest sequence similarity with IP₃R type 2 (IP₃R2) and Ryanodine receptor type 3 (RyR3) which were further used for global sequence similarity and identity analysis by the SIAS server (<http://imed.med.ucm.es/Tools/sias.html>). Such analysis revealed that the putative IP₃R from *Trypanosoma brucei* had a global sequence identity and similarity of 11.75% and 49.95% respectively with IP₃R2 and 5.33% and 9.3% respectively with RyR3.

Domain analysis was conducted using the InterProScan server (<http://www.ebi.ac.uk/Tools/pfa/iprscan/>) and transmembrane domains were determined by the TMHMM server (<http://www.cbs.dtu.dk/services/TMHMM/>). Results are presented relative to scale of the full-length amino acid (AA) sequence of the protein.

(C) Analysis of transmembrane domains of putative IP₃R from *Trypanosoma brucei* (left) and mouse IP₃R2 (right) using the TMHMM server (<http://www.cbs.dtu.dk/services/TMHMM/>).

(D) Hydrophobicity analysis of the transmembrane domain of putative IP₃R from *Trypanosoma brucei*.

Hydrophobicity plots reveal the presence of 5 transmembrane regions compared to 6 that are present in the mouse IP₃R2 (see Panel C (right)).

(E) Clustal W2.0.12 alignment of the amino acid sequence between transmembrane domains 4 and 5 of putative IP₃R from *Trypanosoma brucei* and the amino acid sequence between transmembrane domains 5 and 6 of mouse IP₃R2 and mouse RyR3. The selectivity filter of mouse IP₃R2 is shown as boxed text.

(F) Modeling of the putative IP₃R-ligand binding suppressor domain of *Trypanosoma brucei* using the EsyPred3D homology-modelling server (<http://www.fundp.ac.be/sciences/biologie/urbm/bioinfo/esypred/>). Structure of the IP₃R-ligand binding suppressor domain of mouse IP₃R type 1 (right), (ITPR1; <http://www.pdb.org/pdb/explore/explore.do?structureId=1XZZ>) (Bosanac *et al.*, 2005) and model of the *Trypanosoma brucei* (Tbg; left) homologous region. Numbers indicate the amino acid residues used.

(G) Modeling of the putative IP₃R-ligand binding core from *Trypanosoma brucei* using the EsyPred3D homology-modelling server (<http://www.fundp.ac.be/sciences/biologie/urbm/bioinfo/esypred/>). Structure of the IP₃R-ligand binding core of mouse IP₃R type 1 (right), (ITPR1; <http://www.pdb.org/pdb/explore/explore.do?structureId=1N4K>) (Bosanac *et al.*, 2002) and model of the *Trypanosoma brucei* (Tbg; left) homologous region. Numbers indicate the amino acid residues used. The arrow indicates the IP₃ binding site.

(H) Evolutionary relationship of the putative IP₃Rs from *Leishmania infantum* and *Trypanosoma brucei*.

Predictions from multiple sequence alignments are shown in a neighbor-joining tree with 1000 bootstrap replicates generated with the PHYLIP software package (<http://mobyli.pasteur.fr/cgi-bin/portal.py>). Sequence representing the mammalian IP₃R was from *Mus musculus* (Mouse IP₃R2 (IP₃R type 2, AB182288). Sequence representing the mammalian Ryanodine receptor was from *Mus musculus* (Mouse RyR3 (Ryanodine receptor type 3, NM_177652). Other metazoa IP₃R sequence was from *Caenorhabditis elegans* (AJ243179) and protozoa IP₃R sequence was from *Paramecium tetraurelia* (CR932323). Evolutionary distances are indicated by the scale bar.